

Control of postharvest gray mold of ‘BRS Nubia’ table grape under cold storage

Controle de mofo cinzento na pós-colheita da uva de mesa ‘BRS Nubia’ sob armazenamento refrigerado

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Highlights:

Gray mold control on ‘BRS Nubia’ table grapes can be achieved using an SO₂ pads.
‘BRS Nubia’, a novel table grape, may be stored for up to 30 days under cold storage.
SO₂ pads do not affect the chemical characteristics of grape berries.

Abstract

The demand for high-quality nutritional products has increased fruit consumption, as grapes, for this reason postharvest techniques are required to prevent losses, to preserve quality, to extend shelf life, and to attend to consumer needs. In this way, the objective of this study was to evaluate strategies to control gray mold caused by *Botrytis cinerea* in ‘BRS Nubia’ grapes during cold storage and shelf life periods. Grape bunches were harvested from a commercial vineyard in Marialva, Parana, Brazil. Grapes were subjected to the following treatments: cold storage at 2 °C (control), cold storage at 2 °C with SO₂-generating pads, cold storage at 2 °C and inoculated with *B. cinerea* suspension, and cold storage at 2 °C with SO₂-generating pads and inoculated with *B. cinerea* suspension. The experiment was conducted in a complete randomized design with five replications per treatment using four bunches per experimental unit. A factorial arrangement (absence/presence of SO₂ pads × absence/presence of *Botrytis* inoculation) was applied. At the end of 30 days of cold storage and 7 days of shelf life (22 °C), gray mold incidence, shattered berries, and physicochemical parameters were evaluated. The gray mold incidence on ‘BRS Nubia’ grapes decreased when SO₂-generating pads were used during cold storage. Berry weight loss was greater in the treatments without SO₂-generating pads after 30 days of cold storage followed by 7 days of shelf life. Berry firmness, soluble solids content (SS), total acidity (TA), SS/TA ratio, and anthocyanins concentration were not negatively affected by SO₂-generating pad treatments. However, a slight increase in the shattered berries percentage was recorded for the SO₂-generating pad treatments. No significant quality loss of ‘BRS Nubia’ grape was evident after 30 days of cold storage followed by 7 days of exposure at room temperature. In this context, SO₂-generating pads can be used to control the gray mold incidence on ‘BRS Nubia’ table grapes during cold storage.

Key words: *Botrytis cinerea*. Fruit quality. Sulfur dioxide.

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Resumo

A demanda por produtos de alta qualidade nutricional tem aumentado o consumo de frutas, como as uvas, por esse motivo, são necessárias a adoção de técnicas pós-colheita para evitar perdas, preservar a qualidade e prolongar a vida útil, além de atender às necessidades do consumidor. Dessa forma, o objetivo deste estudo foi avaliar estratégias para o controle do mofo cinzento causado por *Botrytis cinerea* em uvas ‘BRS Nubia’ durante o armazenamento refrigerado e a vida de prateleira. Cachos da uva ‘BRS Nubia’ foram colhidos em um vinhedo comercial localizado em Marialva, Paraná, Brasil. As uvas foram submetidas aos seguintes tratamentos: armazenamento refrigerado à 2 °C (controle); armazenamento refrigerado à 2 °C com folha geradora de SO₂; armazenamento refrigerado à 2 °C, inoculado com suspensão de *B. cinerea*; e armazenamento refrigerado à 2 °C com folha geradora de SO₂, inoculado com suspensão de *B. cinerea*; empregando o delineamento inteiramente casualizado, arranjado em esquema fatorial (ausência / presença de SO₂ x ausência / presença de *Botrytis*), com cinco repetições e quatro cachos por repetição. Após 30 dias de armazenamento refrigerado e sete dias sob temperatura ambiente (22 °C), avaliou-se a incidência de mofo cinzento, degrana e características físico-química das bagas. A incidência de mofo cinzento em uvas ‘BRS Nubia’ diminuiu com o uso de folhas geradoras de SO₂ durante o armazenamento refrigerado. A perda de massa das bagas foi maior nos tratamentos sem folhas geradoras de SO₂ após 30 dias de armazenamento refrigerado e 7 dias de prateleira. A firmeza das bagas, a concentração de sólidos solúveis (SS), acidez total (TA), SS/TA e antocianinas totais não foram afetadas pelos tratamentos com folhas geradoras de SO₂. No entanto, houve aumento na porcentagem de degrana das bagas nesses tratamentos. Não foi observada perda significativa de qualidade das uvas ‘BRS Nubia’ mantidas em temperatura ambiente por um período de 7 dias, após os 30 dias de armazenamento refrigerado.

Palavras-chave: *Botrytis cinerea*. Dióxido de enxofre. Qualidade do fruto.

In response to the fresh market, where consumers demand products of high quality, the maintenance and improvement of fruit quality during postharvest life have become increasingly important in recent years. Among these fruits, grapes have been recognized to have a wide range of compounds that improve health, such as phenolic compounds that neutralize free radicals (Colombo et al., 2020).

‘BRS Nubia’ is a seeded hybrid table grape released by Embrapa Grape and Wine, Brazil, with black color and neutral flavor. It is a highly productive cultivar that grows large and crunchy berries (Maia et al., 2013). This grape cultivar loads large (~0.7 kg) and slightly compact bunches, demanding thinning techniques to reduce compactness (Silvestre et al., 2017). Compact bunches impair uniform fungicide spraying, which favors the growth of fungal spores or may become latent in the innermost area of the bunch developing at postharvest. Adapted to cultivation under subtropical areas with mild winters, ‘BRS Nubia’

can grow under a two-crop one-year system and is a strong candidate for overseas markets. However, in this intensive production system, the harvest period may coincide with periods of climatic conditions favorable to the development of postharvest fungal diseases, which can limit the long-distance transportation of fresh grapes (Youssef et al., 2015).

Among the table grapes postharvest diseases, the typical necrotroph attacking grapevines is *Botrytis cinerea* Pers. ex Fr., which can infect all green plant organs but is best known to cause gray mold in grapes and many other plant species (Carisse & Van Der Heyden, 2015; Hashim, Youssef, & Abd-Elsalam, 2019). Infections by *B. cinerea* can occur before harvest but can remain latent up to storage, when the pathogen takes advantage of disease development from higher relative humidity and low temperatures that slow down host defenses. The traditional control of gray mold infection consists of field application of synthetic fungicides during the crop growing cycle (Romanazzi & Feliziani, 2014).

Several families of synthetic fungicides are available to control gray mold infections (Romanazzi & Feliziani, 2014). However, the control of gray mold is very difficult through synthetic fungicides, and due to safety reasons, they have been banished in many countries (Youssef et al., 2015). Thus, some environmentally friendly strategies are recommended to control gray mold in table grapes during postharvest, such as treatments with salt solutions (Youssef & Roberto, 2014), SO₂-generating pads, and cold storage (Ahmed et al., 2019; Chaves et al., 2019; Youssef et al., 2015).

However, aiming to maintain the quality of the harvested product during transport, the selection of SO₂-generating pads should be judicious, avoiding causing toxic effects to the consumer or changes in the sensory profile of the grapes. Additionally, the level of active ingredients must be appropriate to avoid damaging the fruit or impairing its flavor. Thus, information regarding 'BRS Nubia' table grapes during cold storage in relation to gray mold development has not yet been explored, and it is a very important issue for long distance and overseas markets. In this context, the objective of this work was to evaluate the control of gray mold of 'BRS Nubia' table grapes using SO₂-generating pads and the effect on bunch quality during cold storage and shelf-life periods.

Bunches of 'BRS Nubia' table grapes were harvested in mid-May 2016 from a two-year-old commercial vineyard in Marialva, state of Parana, South Brazil (23°29'S, 51°47'W, altitude 570 m). The vines were grafted on 'IAC 766 Campinas' rootstock, trained on overhead trellis with 18% plastic mesh covered, and spaced at a distance of 2.5 × 9.0 m apart.

Botrytis cinerea was isolated from infected grapes showing typical gray mold symptoms, purified and identified morphologically and molecularly, as described by Youssef and Roberto (2014). The isolates were maintained on PDA slants and stored at 4 °C for further use. Fungal mycelia

were harvested from 2-week-old PDA cultures of *B. cinerea* grown at 23±1 °C. A volume of 5 mL of sterile water containing 0.05% (v/v) Tween 80 was added to a Petri plate culture. The mycelium was gently dislodged from the surface with a sterile glass rod and crushed with a mini processor. The mycelium suspensions were diluted with sterile water, and the concentration was determined with a haemocytometer. Further dilutions with sterile water were made to obtain the desired concentration. A *B. cinerea* suspension (10⁶ mycelium sections per mL) was used for grape inoculation.

Bunches of 'BRS Nubia' table grapes of approximately 0.720 kg, 14.1 °Brix, 0.8% tartaric acid and 14.8 mg g⁻¹ anthocyanins were harvested in the morning and selected manually to remove the damaged berries. Bunches were submitted to the following treatments: cold storage at 2 °C (control); cold storage at 2 °C with an SO₂-generating pad; cold storage at 2 °C and inoculated with a *B. cinerea* mycelium suspension; and cold storage at 2 °C with an SO₂-generating pad and inoculated with a *B. cinerea* mycelium suspension. The bunches that were inoculated with *B. cinerea* mycelium suspension were previously disinfested according to Chaves et al. (2019). For SO₂ treatments, one generator pad measuring 13 cm × 23 cm per box provided with fast and slow release phases of sodium metabisulfite (Na₂S₂O₅), 98% (Matesa®, Grapeguard, Chile) was used during the cold storage period and subsequently removed at the beginning of shelf life. The experiment was conducted in a complete randomized design with five replications per treatment using four bunches per experimental unit. A factorial arrangement (absence/presence of SO₂ pads × absence/presence of *Botrytis* inoculation) was applied.

The inoculation was carried out by spraying the mycelium suspension, and the bunches were left to dry at room temperature (22±2 °C) for 1 h and placed into carton boxes (13 × 30 × 40 cm). The boxes were covered with colorless plastic bags and stored in cold chambers for 30 days at 2±1 °C and

high RH, followed by a 7-day period of shelf life at 22 ± 2 °C. By the end of 30-day cold storage and 7-day shelf-life periods, the following variables were evaluated for grape quality measurements:

1. *Gray mold incidence and shattered berries:*

The incidence of gray mold on bunches was measured according to the following formula: disease incidence (%) = (number of decayed berries per bunch/total number of berries per bunch) \times 100 (Youssef & Roberto, 2014). Additionally, we evaluated the percentage of shattered berries per plot as follows: shattered berries (%) = (number of shattered berries per plot/total number of berries per plot) \times 100.

2. *Physicochemical analysis:* Weight loss (%) was calculated as a percentage of bunch weight at the beginning and end of the cold storage/shelf-life period. The difference as a percentage from the initial weight was calculated [weight loss (%) = (initial weight - weight at examined date/initial weight) \times 100].

Berry firmness (N) analysis was performed for each period of evaluation with a texture analyzer TA. XT Plus. The measurements were taken from the equatorial position of 10 berries with pedicels per plot. Each berry was placed on the base of the equipment and compressed using a cylindrical probe with a diameter of 35 mm parallel to the base. A constant force of 0.1 N at a speed of 1.0 mm s^{-1} was applied to promote the cracking of the sample.

The chemical parameters of bunches were assessed by determining the soluble solids (SS, °Brix), total acidity (TA, % of tartaric acid), SS/TA ratio, and total anthocyanin content in berry skin [mg of anthocyanins (as malvidin-3-glucoside) per gram of skin]. To perform the chemical analysis, five berries were collected from each box, totaling 15 berries per plot.

The data were subjected to analysis of variance (ANOVA), and the means were compared by Tukey's test at the $p\leq 0.05$ level.

No significant interaction ($p\leq 0.05$) was found between the tested factors; however, isolated effects of SO_2 -generating pads and *B. cinerea* inoculation on some parameters were observed. For the gray mold symptoms in both conditions, at 30-day period of cold storage at 2 °C and at the end of 7-day of shelf life, the factor *B. cinerea* inoculation was evident in relation to non-inoculated treatments.

After the 30-day period of cold storage at 2 °C, gray mold symptoms were observed in all treatments (Table 1). However, in the treatments without *B. cinerea* inoculation, the gray mold incidence was significantly lower (4.40%) than that with inoculation (11.05%), which shows the effectiveness of the use of mycelium suspension as an inoculum source. We also observed a significant effect of the SO_2 -generating pad on gray mold control. The lowest gray mold incidence (5.08%) was recorded in grapes subjected to cold storage using SO_2 -generating pads, whereas the treatments without SO_2 -generating pads reached a mid-incidence of gray mold (9.73%).

At the end of the 7-day shelf life period, the incidence of gray mold was higher than that of the 30-day cold storage period in all treatments (Table 1). Again, the *B. cinerea* inoculation in bunches showed a significant effect on disease severity compared to non-inoculated bunches. In contrast, no significant effect was found for the use of SO_2 -generating pads on gray mold control under shelf-life conditions.

The use of SO_2 -generating pads contributed to an increase in shattered berries after 30 days of cold storage and at the 7-day shelf life period, reaching 5.96% and 10.36%, respectively. On the other hand, no significant effect of *B. cinerea* inoculation was observed on shattered berries.

Table 1**Gray mold and shattered berry incidence in 'BRS Nubia' table grapes submitted to different postharvest treatments at 30 days of cold storage (CS) and 30 days of cold storage with 7 days of shelf life**

Treatments	Gray Mold (%)	Shattered berries (%)
	30-day of cold storage	
(-) SO ₂ pad	9.73a	2.87b
(+) SO ₂ pad	5.08b	5.96a
(-) <i>Botrytis</i> inoc.	4.40B	4.55
(+) <i>Botrytis</i> inoc.	11.05A	4.28
CV (%)	23.87	60.61
	30-day of CS + 7 days of shelf-life	
(-) SO ₂ pad	13.45	5.31b
(+) SO ₂ pad	11.39	10.36a
(-) <i>Botrytis</i> inoc.	5.21B	8.51
(+) <i>Botrytis</i> inoc.	18.99A	7.15
CV (%)	38.23	62.87

(-) absence and (+) presence. Means followed by different letters, lowercase letters for SO₂ pad and capital letters for *Botrytis* inoculation, differ significantly according to Tukey's test ($p \leq 0.05$).

In relation to weight loss, no significant effects were recorded among the treatments at 30 days of cold storage. However, a significant reduction was observed for the SO₂-generating pad treatment at the end of shelf life. In general, regardless of treatment, the weight loss ranged from 3.28 to 3.97% across the evaluated period (Table 2). Regarding berry firmness, Tukey's test ($p \leq 0.05$) found no differences between the treatments in both periods of evaluation (Table 2).

No significant effects of SO₂-generating pads or *B. cinerea* inoculation were found on the chemical parameters of the berries in the first evaluation at 30 days of cold storage. On the other hand, a significant effect of *B. cinerea* inoculation on SS, TA and SS/TA ratio parameters was observed at the end of 7 days of shelf life. Berries from non-inoculated bunches showed higher SS content and SS/TA ratio and lower TA than inoculated bunches. Interestingly, a reduction of ~50% in the total anthocyanin contents was also observed when the periods were compared (Table 2).

Gray mold is the most important disease occurring postharvest in grapes, and it is responsible

for major quality losses in grape bunches and berries in the main grape-growing areas of the world. In this trial, the percentage of gray mold incidence was evaluated under natural and artificial conditions, since field treatments are not efficient, and some infections that occur in the field remain quiescent during the growing season and develop after harvest (Romanazzi, Smilanick, Feliziani, & Droby, 2016).

B. cinerea strains resistant to fungicides are very common and emphasize the need for an accurate adoption of Fungicide Resistance Action Committee (FRAC) guidelines, suggesting a limited number of applications per year. Furthermore, the traditional control of postharvest infection in cold-stored table grapes can be achieved by repeated fumigation with SO₂ or using SO₂-generating pads combined with polyethylene-lined grape containers (Youssef et al., 2015).

At 30 days of cold storage, the gray mold incidence under natural conditions (non-inoculated) was significantly lower than that under *B. cinerea* inoculation treatments, approximately 4 and 11%, respectively. However, previous studies reported higher gray mold incidence on 'BRS Vitoria'

table grapes grown in the same area, 20 and 40% under natural and artificial (inoculated) conditions, respectively (Youssef et al., 2015). Thus, this result may be associated with cultivar tolerance, climate conditions along with crop season, and field

management. Regardless of the incidence level, it is crucial to control this disease, especially when grapes have to be stored in cold chambers before commercialization.

Table 2

Weight loss (WL), berry firmness (BF), soluble solids (SS), titratable acidity (TA), SS/TA ratio, and total anthocyanin content (ACNs) of ‘BRS Nubia’ table grapes submitted to different postharvest treatments at 30 days of cold storage and 30 days of cold storage with 7 days of shelf life

Treatments	WL (%)	BF (N)	SS (°Brix)	TA (%)	SS/TA	ACNs (mg g ⁻¹)
30-day of cold storage						
(-) SO ₂ pad	3.34	8.48	14.11	0.58	24.64	11.85
(+) SO ₂ pad	3.53	8.56	14.20	0.57	25.32	13.83
(-) <i>Botrytis</i> inoc.	3.59	8.54	14.30	0.58	25.18	13.28
(+) <i>Botrytis</i> inoc.	3.28	8.50	14.01	0.57	24.79	12.40
CV (%)	9.65	8.29	4.23	10.83	11.89	24.60
30-day of cold storage + 7 days of shelf-life						
(-) SO ₂ pad	3.97a	7.74	14.47	0.74	20.04	6.20
(+) SO ₂ pad	3.36b	7.90	14.46	0.74	20.31	7.28
(-) <i>Botrytis</i> inoc.	3.46	7.74	14.90A	0.70B	21.56A	7.29
(+) <i>Botrytis</i> inoc.	3.86	7.91	14.03B	0.78A	18.79B	6.19
CV (%)	16.59	6.13	5.08	8.28	10.92	18.95

(-) absence and (+) presence. Means followed by different letters, lowercase letters for SO₂ pad and capital letters for *Botrytis* inoculation, differ significantly according to Tukey's test ($p \leq 0.05$).

The use of SO₂-generating pads was shown to be a good strategy to control gray mold when ‘BRS Nubia’ table grapes are submitted to cold storage, with satisfactory results at the end of 30 days. These results agree with those reported by Ahmed et al. (2019), Chaves et al. (2019), and Youssef et al. (2015) for some table grape cultivars. Depending on grape cultivar, the results are even better, as those obtained for ‘Red Globe’ grapes, in which SO₂-generating pads can control the disease up to a 4-month period under cold storage (Ozkaya, Dundar, & Özdemir, 2008). However, in this specific case, the flavor of grapes declined slightly and remained acceptable until the 3-month period of storage.

SO₂-generating pads did not show a significant effect on controlling the gray mold incidence at 7 days of shelf life. A possible explanation for

this might be that the residual effect of SO₂ in grape berries was very low and did not suppress pathogen development under shelf-life conditions. These results were expected because bunches were exposed to favorable conditions for disease development. In addition, the carton boxes filled with grape bunches remained opened along the shelf-life period, favoring SO₂ gas dispersal.

Regarding the physical parameters of berries, the weight loss ranged from 3.28 to 3.97%, and a low reduction in berry firmness was observed during the evaluated periods (Table 2). ‘BRS Nubia’ table grape has a large berry, approximately 11 g (Silvestre et al., 2017); for this reason, the low weight loss and reduction in berry firmness appear not to be commercially significant.

In contrast, the chemical characteristics of berries, such as SS, TA, SS/TA ratio, and anthocyanin content, were not affected by SO₂-generating pads and *B. cinerea* inoculation over the cold storage period. However, at the end of shelf life, *B. cinerea* inoculation contributed to decreasing SS content. It seems possible that the greater severity of gray mold on the inoculated bunches affected the berry quality. Nevertheless, 'BRS Nubia' table grapes harvested in off-season crops showed adequate SS content for domestic and international markets (Table 2), which requires a baseline of 14 °Brix or SS/TA ratio ≥ 20 (Ministério da Agricultura, Pecuária e Abastecimento [MAPA], 2002; United Nations Economic Commission for Europe [UNECE], 2017).

Turning now to the experimental effect of SO₂-generating pads on berry characteristics, such as SS, TA, SS/TA ratio, and anthocyanins, no significant differences were recorded among them. This result reinforces that SO₂-generating pads decreasing gray mold on 'BRS Nubia' table grapes do not alter their chemical characteristics. In this context, 'BRS Nubia' table grapes were shown to be non-sensitive to the amount of SO₂ gas released by the evaluated pads, except for the percentage of shattered berries. Similarly, the SO₂-generating pad did not affect these parameters in 'BRS Isis' and 'BRS Vitoria' seedless grapes (Ahmed et al., 2019; Youssef et al., 2015). However, at high concentrations, this compound can cause bleaching or premature stem browning and may also damage the fruits, resulting in unwanted conditions.

At the end of shelf life, a decrease in total anthocyanin content of approximately 50% was observed in comparison with the end of cold storage, which may have occurred because grapes were exposed to higher temperatures (22±2 °C) than those of cold storage. Similar results were recorded in 'BRS Vitoria' table grapes at the end of the shelf-life period (Colombo et al., 2018). Anthocyanins are usually degraded when fruit are exposed to high temperatures and light conditions. The anthocyanins

were relatively sensitive in açai (*Euterpe oleracea*) fruit, where half of their content was degraded within 48 hours after harvest when fruit were kept at 30 °C (Rogez, Akwie, Moura, & Larondelle, 2012).

In summary, regarding *B. cinerea* inoculation, it was possible to verify disease development at the end of cold storage and shelf life. The growth of gray mold on inoculated bunches impairs the bunch appearance, modulating the chemical parameters of the berries at the end of shelf life and depreciating the product for the domestic and international markets. A decrease in gray mold incidence was proven when the SO₂-generating pads on 'BRS Nubia' table grapes were used during cold storage. Taken together, 'BRS Nubia' table grapes can be stored for up to 30 days under cold storage without significant changes in their chemical parameters.

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