Molecular diagnosis and risk factors of canine ehrlichiosis in the municipality of Itabuna-Bahia, Brazil

Diagnóstico molecular e fatores de risco da erliquiose canina no município de Itabuna-Bahia

Jeane Martinha dos Anjos Cordeiro¹; Paula Elisa Brandão Guedes²; Alexandre Dias Munhoz³; Fabiana Lessa Silva³*

Highlights:

Dogs from Itabuna-Bahia have a high infection level by *Ehrlichia canis*. Contact with other dogs is an important factor for the dissemination of ehrlichiosis. Males had lower risk of infection with *E. canis*.

Abstract

Ehrlichiosis is an emerging zoonosis worldwide and has had several adverse effects on public health. Canine monocytic ehrlichiosis (CME), caused by Ehrlichia canis, has the tick Rhipicephalus sanguineus as the vector. The main clinical signs in affected dogs are fever, apathy, anorexia, weight loss, and neurological signs. The diagnosis is made through the association of clinical signs with parasitological. serological, and molecular tests. The aim of this study was to evaluate the occurrence of E. canis infection in dogs from the city of Itabuna-Bahia, as well as to identify the risk factors related to infection. For this, 405 dogs from the Center for Zoonoses Control (CCZ), non-governmental organizations (NGOs), and dogs domiciled and semi-domiciled in the city of Itabuna, southern Bahia, were evaluated. After initial physical evaluation of the dogs, blood samples were collected by venipuncture for subsequent DNA extraction and E. canis testing using the nested Polymerase Chain Reaction (nested-PCR) technique. In addition, an epidemiological questionnaire that included questions related to the animals was administered to the dog owners to identify the risk factors for exposure to the etiological agent and to the vector. Approximately 17% of the dogs in the municipality of Itabuna-Bahia tested positive for E. canis by nested-PCR, a result higher than that found in other studies conducted in the same municipality. Among the factors associated with *E. canis* infection, contact with other dogs (p = 0.0226) was an important factor for the dissemination of CME, since dogs are reported to be reservoirs of E. canis. Male dogs (p = 0.0016) presented lower risk for *E. canis* infection. Other studies, however, describe no association between animal gender and infection by E. canis. Preventive measures to reduce exposure to the vector of ehrlichiosis are necessary.

Key words: Canis familiaris. Ehrlichia canis. Nested-PCR. Zoonosis. Vector-borne disease.

Received: Mar. 29, 2019 - Approved: Aug. 07, 2019

¹ Discente de Doutorado, Curso de Pós-Graduação em Ciência Animal, Universidade Estadual de Santa Cruz, UESC, BA, Brasil. E-mail: jeane.martinha@gmail.com

² Pós-Doutoranda PNPD, Programa de Pós-Graduação em Ciência Animal, UESC, BA, Brasil. E-mail: paulaebg@gmail.com

³ Prof^{as.} Pleno, Departamento de Ciências Agrárias e Ambientais, UESC, BA, Brasil. E-mail: munhoz@uesc.br; fabiana.lessa@ gmail.com

^{*} Author for correspondence

Resumo

A erliquiose é uma zoonose emergente em todo o mundo e tem acarretado diversos transtornos para a saúde pública. A erliquiose monocítica canina (EMC), causada pela Ehrlichia canis, tem como vetor o carrapato Rhipicephalus sanguineus. Os principais sinais clínicos nos cães afetados são febre, apatia, anorexia, perda de peso e sinais neurológicos. O diagnóstico é feito através da associação dos sinais clínicos, exames parasitológicos, sorológicos e moleculares. Objetivou-se através deste estudo avaliar a ocorrência de infecção por E. canis em cães do município de Itabuna-Bahia, bem como identificar os fatores de risco relacionados à infecção. Para tanto, foram avaliados 405 cães provenientes do Centro de Controle de Zoonoses (CCZ), Organizações não governamentais (ONGs), cães domiciliados e semi-domiciliados da cidade de Itabuna-Bahia. Após avaliação física inicial dos cães, procedeu-se em seguida a coleta de amostras de sangue por punção venosa, para posterior extração de DNA e pesquisa de E. canis pela técnica de nested-Reação em Cadeia de Polimerase (nested-PCR). Adicionalmente, um questionário epidemiológico foi aplicado junto aos responsáveis, no qual constavam questões relacionadas aos animais, com a finalidade de identificar os fatores de risco de exposição destes ao agente etiológico e ao vetor. Em suma, este estudo mostrou que aproximadamente 17% dos cães do município de Itabuna-Bahia foram positivos para E. canis pela nested-PCR, resultado superior ao encontrado em outros estudos realizados no mesmo município. Dos fatores associados à infecção por E. canis, foi significativo o contato com outros cães (p=0,0226), um fator importante para a disseminação da EMC, pois os cães são relatados como reservatórios da E. canis. O gênero macho (p=0,0016) apresentou menor risco para a infecção por E. canis. Outros estudos, no entanto, descrevem que não há nenhuma associação entre o gênero do animal e a infecção por E. canis. Adverte-se a população sobre a necessidade de medidas profiláticas para diminuir a exposição ao vetor da erliquiose. Palavras-chave: Canis familiaris. Ehrlichia canis. Nested-PCR. Zoonose. Doenças transmitidas por vetores.

Introduction

Ehrlichiosis is a zoonosis that affects many species of animals, such as domesticated animals including dogs, cats, horses, ruminants, as well as wild animals and humans (Perez, Bodor, Zhang, Xiong, & Rikihisa, 2006; Otranto, Dantas-Torres, & Breitschwerdt, 2009; Isola, Cardioli, & Nakage, 2012; Vieira et al., 2013; Pritt et al., 2017). The literature describes three diseases in dogs that are caused by different species of Ehrlichia: canine monocytic ehrlichiosis, caused by *Ehrlichia canis*; granulocytic ehrlichiosis, caused by the agents *E. ewingii* e *Anaplasma phagocytophilum*; and thrombocytic ehrlichiosis, caused by por *A. platys* (Dagnone, Morais, & Vidotto, 2001).

Canine monocytic ehrlichiosis (CME), also known as tropical canine pancytopenia, canine hemorrhagic fever, or tick typhus (Vieira et al., 2013; I. P. M. Silva, 2015), is an endemic disease in tropical and subtropical regions, with reports in temperate regions. The geographical distribution of CME is related to the presence of the vector, the tick *Rhipicephalus sanguineus* (Vieira et al., 2013; Krawczak et al., 2015).

In Brazil, CME is present almost everywhere, since *R. sanguineus* is well adapted to tropical and subtropical regions (Accetta, 2008; Vieira et al., 2011; Ribeiro et al., 2017), with prevalence ranging from 7% to 60% (Dagnone et al., 2001; Fonseca, Hirsch, & Guimarães, 2013; I. P. M. Silva, 2015).

CME can have three clinical stages: acute, subclinical, and chronic. In the acute stage of the disease, the dog presents with fever, depression, and anorexia. In the subclinical phase, the animal presents predominantly with hematological alterations such as thrombocytopenia, leukopenia, and anemia. In the chronic stage, the same signs as in the acute phase are present, in addition to bone marrow hypoplasia and some neurological signs such as ataxia, neuromotor dysfunction, and central or peripheral vestibular dysfunction (Morais, Hoskins, Almosny, & Labarthe, 2004; Moretti, Silva, Ribeiro, Paes, & Langoni, 2006; Saito, 2009; Isola et al., 2012; I. P. M. Silva, 2015).

The diagnosis of CME is based on clinical signs, in conjunction with the results of parasitological, serological, and molecular tests (Carvalho, Wenceslau, Carlos, & Alburquerque, 2008). The polymerase chain reaction (PCR) enables a more accurate diagnosis of CME, with good results in both acute and chronic phase animals (Iqbal & Rikihisa, 1994). Besides having high sensitivity and specificity, PCR allows differentiation between species of the genus Ehrlichia, unlike other diagnostic techniques. Furthermore, due to its high sensitivity, it is able to identify a small amount of parasite DNA, equivalent to only five copies of *E. canis* (Ueno et al., 2009).

The increase in the number of cases of ehrlichiosis in recent years has motivated further research, due to both the zoonotic character of this disease and the high morbidity and mortality rates in dogs (Dagnone et al., 2001; Menezes, Souza, Teixeira, & Guimarães, 2008; Guedes et al., 2015). Thus, the aim of the present study was to evaluate the occurrence of *E. canis* infection in dogs of Itabuna-Bahia, Brazil, as well as to determine the factors associated with exposure to the agent.

Material and Methods

Study area and animals

The study was conducted in the municipality of Itabuna (Latitude 14°47'08" South; Longitude 39°16'49" West), located in the Southern Bahia Region, Ilhéus-Itabuna Microregion, State of Bahia, Brazil (1). The city is about 426 kilometers from the state capital and is part of the Atlantic Forest biome. The municipality has an estimated population of 219,680,000 inhabitants, a total area of 401,028 km² and a population density of 473,50 inhabitants/km². The average rainfall is 1419 mm per year (Instituto Brasileiro de Geografia e Estatística [IBGE], 2016).



Figure 1. Location of the municipality of Itabuna in Bahia. Fonte: http://www.skyscrapercity.com/showthread.php?t=463643

The study included 405 dogs from an estimated population of 32,839, with age ranging from four months to 15 years, of both sexes, which were sampled for convenience. The dogs were domiciled, without free street access (162 animals), or semi-domiciled, with some street access (173 animals). Some came from the Zoonosis Control Center (ZCC) (22 dogs) and non-governmental organizations (NGOs) (48 dogs).

Sample collection was performed from March to June 2016. Prior to the inclusion of the animals in the study, the owners or guardians of the dogs were informed about the research, and after agreement, signed a free and informed consent. This study was approved by the Animal Use Ethics Committee (CEUA) of the Santa Cruz State University located in the municipality of Ilhéus, southern Bahia, Brazil (Protocol 034/15).

History, clinical examination, and epidemiological data collection

An epidemiological questionnaire was administered to dog owners and/or guardians. Questions related to animals (sex, age, presence of ticks, and contact with other dogs) were asked, in order to identify the risk factors associated with infection. Anamnesis and physical examination of all dogs included in the study were performed, in which mucosa staining, hydration, lymph node size, tick parasitism, presence of petechiae, and ecchymoses in the skin and mucosa were verified, in order to investigate the occurrence of physical changes consistent with the disease.

Biological samples collection

After physical restraint, 5 mL of blood was collected from each dog through jugular or cephalic venipuncture using sterile syringes and needles. The samples were stored in tubes with ethylenediamine tetraacetic acid anticoagulant (EDTA) and stored in a recyclable ice pack. Subsequently, the leukocyte cap, which corresponds to the layer where most of the leukocytes and platelets are concentrated was collected via centrifugation. and separation by density gradient. The materials were then stored in Eppendorf-like tubes at -20 °C until DNA extraction.

DNA extraction and polymerase chain reaction

DNA was extracted from the leukocyte coat using the Easy-DNA kit (Invitrogen®), following the manufacturer's instructions. The extracted DNA was quantified using Nanodrop and then stored at -20 °C. For E. canis DNA amplification, a nested PCR was performed. In the first reaction, forward ECC (5'-AGAACGAACGCTGGCGGCAAGC-3 ') and reverse ECB (5'-CGTATTACCGCGGCTGCTGGCA-3') primers were used, which amplify part *Ehrlichia* spp. 16SrRNA gene. In of the the second reaction, the forward ECAN '-CAATTATTATAGCCTCTGGCTATAGGA-3') and reverse HE3 (5'TATAGGTACCGTCATTATCTTCCCTAT-3') primers were used, according to the methodology described by Murphy, Ewing, Whitworth, Fox and Kocan (1998), to give a final product of 396 base pairs.

Each reaction mixture was prepared to a final volume of 25 µL and contained 0.2 mM of each dNTP, 1.5 mM MgCl., 50 mM KCl, 10 mM Tris-HCl, pH 9.0, 0.2 µM of each primer, 2U TaqDNA polymerase (Invitrogen®), and 6µL of purified DNA. Nested-PCR reactions used 3 µL of amplified products. The first reaction amplification protocol consisted of initial denaturation for 3 minutes at 94 °C, followed by 35 cycles of denaturation at 94 °C for 1 minute, annealing at 68 °C for 2 minutes, extension at 72 °C for 2 minutes, and a final extension 72 °C for 7 minutes (Carvalho et al., 2008). For the second reaction, initial denaturation was performed for 3 minutes at 94 °C, followed by 35 cycles of denaturation at 94 °C for 1 minute, annealing at 58 °C for 2 minutes, extension at 72 °C for 1.5

minutes, with final extension at 72 °C for 7 minutes (Carvalho et al., 2008). Nested-PCR products were submitted to 1.5% agarose gel electrophoresis containing Sybrgreen (SYBR Safe DNA gel stain - Invitrogen). Positive controls were samples positive for *E. canis* (Guedes et al., 2015). Ultrapure water was used as negative control. Bands were verified using an ultraviolet transilluminator, followed by photocumentation on the Locus Biotechnology L-Pix Transilluminator.

Statistical analysis

The analysis of the results was performed based on the association between risk factors and the results obtained in nested-PCR. The data obtained from the physical inspection of the animals were also analyzed. Data were analyzed using the Chi-square test or Fisher's exact test with the aid of Epi-Info 7.2.0.1. Bivariate analysis and then multivariate analysis were performed using unconditional logistic regression to identify risk factors related to *E. canis* infection. In the bivariate analysis, all independent variables were crossed with the dependent variables (positive or negative for *E. canis*), selected, and subjected to Spearman correlation to verify multicollinearity (p > 0.8).

Results

Of the 405 dogs evaluated, 67 dogs (16.54%) tested positive for *E. canis*. The physical changes observed in the animals are shown in Figure 2. The presence of petechiae/ecchymoses (p = 0.016) in the skin and mucosa was more frequent in animals positive for *E. canis* (Table 1). Among the factors associated with *E. canis* infection, male sex (p = 0.0016) was a factor associated with lower risk of infection, while contact with other dogs (p = 0.0226) was a risk factor (Tables 2 and 3). There was no collinearity between the variables (p > 0.8).

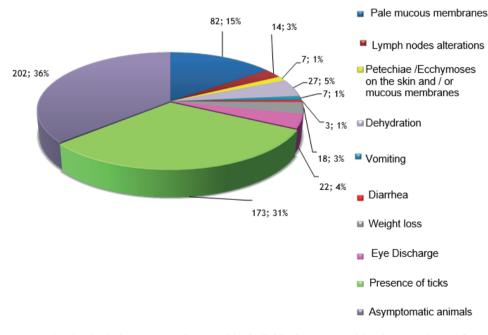


Figure 2. Physical changes consistent with ehrlichiosis presented by dogs evaluated from Itabuna-Bahia.

Variable	1	N	Positive dogs	Prevalence	OR	95% CI	P-value
Pale mucous membranes	Yes No	82 323	12 55	14,63% 17,02%	0,83 1*	0,42-1,64	0,72
Lymph nodes alterations	Yes No	14 391	1 66	7,14% 16,87%	0,37 1*	0,04-2,94	0,55
Petechiae/ Ecchymoses	Yes No	7 398	4 63	57,14% 15,82%	7,08 1*	1,54-32,44	0,016
Dehydration	Yes No	27 378	5 62	18,51% 16,40%	1,15 1*	0,42-3,17	0,98
Vomiting	Yes No	7 398	2 65	28,57% 16,33%	2,04 1*	0,38-10,79	0,72
Diarrhea	Yes No	3 402	0 67	- 16,66%	- 1*	-	1,00**

Table 1

Findings observed on physical examination of dogs from the municipality of Itabuna-Bahia positive for *E. canis* in nested-PCR

*Reference

**Fisher's test.

Table 2 Bivariate analysis of the variables related to *E. canis* infection in positive dogs from Itabuna-Bahia

Variable	Ν		Positive dogs	Prevalence	OR	95% CI	P-value
Tick Presence	Yes No	173 232	29 38	16, 76% 16,37%	1,02 1*	0,60-1,74	1,00
Age (years)	<5 years >5 years	272 133	46 21	16,91% 15,79%	0,98 1*	0,55-1,76	1,00
Contact with other dogs	Yes No	198 207	41 26	20,70% 12,56%	1,81 1*	1,06-3,10	0,038
Gender	Female Male	213 192	47 20	22,06% 10,41%	1* 0,41	0,23-0,72	0,003

*Reference.

Table 3

Multivariate analysis of the association between dogs positive for *E. canis* and gender and contact with other dogs, in the municipality of Itabuna-Bahia

Variable	Odds Ration	Confidence Interval 95%	<i>P</i> -value
Contact with other dogs	1,88	1,09-3,23	0,0226
Male gender	0,40	0,22-0,70	0,0016

Likelihood= 0,004.

Discussion

The frequency of infection found in this study was higher than that observed by Carvalho et al. (2008), who reported 7.8% prevalence of E. canis infection through nested-PCR using the same primers, in dogs from the cities of Ilhéus and Itabuna. This higher percentage of cases in the municipality of Itabuna verified in this study may be due to the fact that the sampling was more significant compared to that of Carvalho et al. (2008), who evaluated only 69 samples from this municipality. Other studies conducted in the state of Bahia found higher prevalence rates of E. canis infection. Using nested-PCR and the same primers, Menezes et al. (2008) and Guedes et al. (2015) reported 33.3% and 25.6% of positive animals in the municipalities of Salvador and Ituberá, respectively.

In addition, most of the animals evaluated came from the ZCC, NGOs, and poor neighborhoods, where a higher frequency of infection was expected, as these locations usually pose a higher risk of dog exposure to the vector tick. This greater exposure occurs due to contact with other dogs, which facilitates the acquisition of ticks, and there is usually no adequate prophylaxis such as vector tick control using acaricides, identification, and treatment of sick animals to decrease the chance of infestation. (Otranto et al., 2010). This can be reinforced by the result obtained in this study, which shows that animals that had contact with other dogs were more likely to become infected than those who did not have this type of contact (Table 3). In a study developed by Azevedo et al. (2011), contact with other dogs was also considered a risk factor for E. canis infection, indicating that this is an important factor for the transmission of CME.

Another factor that may influence the variation between the results is the phase of the disease in which the animal is at the moment the diagnosis is made. In the acute phase of EMC, bacteria are detected in the circulating blood, while in the subclinical or chronic phases, spleen sequestration of infected cells may occur (Faria et al., 2010; Harrus & Waner, 2011). Thus, PCR from blood samples has better sensitivity when the animals are in the acute phase of the disease. During the physical evaluation of the dogs in this study, 202 of the 405 animals did not manifest clinical signs characteristic of CME (Figure 2), and are therefore not in the acute phase of the disease. However, 23 of these 202 animals were positive for E. canis, reinforcing the fact that animals infected with E. canis may have subclinical infection. Animals may remain infected for years without showing clinical signs of the disease. Some authors, however, reported the occurrence of slight hematological changes such as thrombocytopenias, in addition to high specific antibody titers (Harrus & Waner, 2011). Immunocompetent dogs usually recover, while immunocompromised dogs progress to the chronic disease phase (I. P. M. Silva, 2015).

In this study, an association was observed between the presence of petechiae and ecchymoses in the skin and/or mucosa and *E. canis* infection. Saito (2009) and Witter et al. (2013) also associated the presence of these changes with infection with this agent. Generally, animals with CME have vascular disorders associated with thrombocytopenia, which cause petechial hemorrhages and bruises in different organs and body regions (Ueno et al., 2009).

The results obtained in this study indicate that males have lower risk of infection with *E. canis*. Other studies, however, reported no association between animal gender and *E. canis* infection (Nakaghi, Machado, Costa, André, & Baldani, 2008; Sousa et al., 2010; Isola et al., 2012; G. C. Silva et al., 2012).

Although some studies have suggested that age and tick parasitism are risk factors for *E. canis* infection, in this study these factors had no significant effects. This lack of association has also been reported by other authors (Carlos, Carvalho, Wenceslau, Almosny, & Albuquerque, 2011; Souza et al., 2010; Ueno et al., 2009; Witter et al., 2013).

Regarding the absence of ticks in most positive animals at the time of collection (Table 2), it is worth noting the possibility that these dogs were parasitized by *R. sanguineus* before the examination. Although there was no statistical significance in this study, dog exposure to R. sanguineus is of extreme importance for the transmission of CME. Infected ticks only transmit the bacteria for a period of 155 days (Lewis Jr., Ristic, Smith, Lincoln, & Stephenson, 1977); however, dogs can remain infected for up to five years after the initial infection (Trapp et al., 2006; Saito, 2009; Isola et al., 2012). Thus, it is necessary for the tick to become infected by performing blood repast on an infected dog and subsequently transmit the infection to a healthy dog (Saito, 2009; Guedes et al., 2015).

Conclusion

This study revealed a high prevalence of dogs positive for *E. canis* in the municipality of Itabuna-Bahia, Brazil. This result warns of the imminent risk of exposure of the human and canine population to this agent, which is a zoonosis and have adverse effects on public health. Veterinarians should always advise the population on the care of their animals so that exposure to *R. sanguineus* is reduced.

Acknowledgments

Research supported by FAPESB and UESC.

References

- Accetta, E. M. T. (2008). Ehrlichia canis e Anaplasma platys em cães (Canis familiaris, Linnaeus, 1758) trombocitopênicos da região dos lagos do Rio de Janeiro. Dissertação mestrado em Medicina Veterinária, Universidade Federal Rural do Rio de Janeiro, Rio de Janeiro, RJ, Brasil.
- Azevedo, S. S., Aguiar, D. M., Aquino, S. F., Orlandelli, R. C., Fernandes, A. R. F., & Uchoa, I. C. P. (2011). Soroprevalência e fatores de risco associados à soropositividade para *Ehrlichia canis* em cães do semiárido da Paraíba. *Brazilian Journal of Veterinary*

Research and Animal Science, *48*(1), 14-18. doi: 10.11606/S1413-95962011000100002

- Carlos, R. S. A., Carvalho, F. S., Wenceslau, A. A., Almosny, N. R. P., & Albuquerque, G. R. (2011). Risk factors and clinical disorders of canine ehrlichiosis in the South of Bahia, Brazil. *Revista Brasileira de Parasitologia Veterinária*, 20(3), 210-214. doi: 10.1590/S1984-29612011000300006
- Carvalho, F. S., Wenceslau, A. A., Carlos, R. S. A., & Albuquerque, G. R. (2008). Epidemiological and molecular study of *Ehrlichia canis* in dogs in Bahia, Brazil. *Genetics and Molecular Research*, 7(3), 657-662. doi: 10.4238/vol7-3gmr468
- Dagnone, A. S., Morais, H. S. A., & Vidotto, O. (2001). Erliquiose nos animais e no homem. Semina: Ciências Agrárias, 22(2), 191-201. doi: 10.5433/1679-0359.2001v22n2p191
- Faria, J. L. M., Dagnone, A. S., Munhoz, T. D., João, C. F., Pereira, W. A. B, Machado, R. Z., & Tinucci-Costa, M. (2010). *Ehrlichia canis* morulae and DNA detection in whole blood and spleen aspiration samples. *Revista Brasileira de Parasitologia Veterinária*, 19(2), 98-102. doi: 10.4322/rbpv.01902006
- Fonseca, J. P., Hirsch, C., & Guimarães, A. M. (2013). Erliquiose monocítica canina: epidemiologia, imunopatogênese e diagnóstico. *PUBVET*, 7(8), 619-706. doi: 10.22256/pubvet.v7n8.1529
- Guedes, P. E. B., Oliveira, T. N. A., Carvalho, F. S., Carlos, R. S. A., Albuquerque, G. R., Munhoz, A. D., & Silva, F. L. (2015). Canine ehrlichiosis: prevalence and epidemiology in northeast Brazil. *Brazilian Journal of Veterinary Parasitology*, 24(2), 115-121. doi: 10.1590/S1984-29612015030
- Harrus, S., & Waner, T. (2011). Diagnosis of canine monocytotropic ehrlichiosis (*Ehrlichia canis*): an overview. *The Veterinary Journal*, 187(3), 292-296. doi: 10.1016/j.tvjl.2010.02.001
- Instituto Brasileiro de Geografia e Estatística. (2016). *Censo demográfico*. Rio de Janeiro: IBGE. Recuperado de https://cidades.ibge.gov.br/brasil/ba/ itabuna/panorama
- Isola, J. G. M. P., Cadioli, F. A., & Nakage, A. P. (2012). Erliquiose canina-Revisão de literatura. *Revista Científica Eletrônica de Medicina Veterinária*, ANO IX(8), 1-11.
- Iqbal, Z., & Rikihisa, Y. (1994). Reisolation of *Ehrlichia* canis from blood and tissues of dogs after doxycycline treatment. Journal Clinical Microbiology, 32(7), 1644-1649.

- Krawczak, F. S., Reis, I. A., Silveira, J. A., Avelar, D. M., Marcelino, A. P., Werneck, G. L., & Paz, G. F. (2015). *Leishmania, Babesia* and *Ehrlichia* in urban pet dogs: co-infection or cross-reaction in serological methods? *Revista da Sociedade Brasileira de Medicina Tropical*, 48(1), 64-68. doi: 10.1590/0037-8682-0291-2014
- Lewis, G. E. Jr., Ristic, M., Smith, R. D., Lincoln, T., & Stephenson, E. H. (1977). The brown dog tick *Rhipicephalus sanguineus* and the dog as experimental hosts of *Ehrlichia canis. American Journal of Veterinary Research*, 38(12), 1953-1955.
- Menezes, S. D. I., Souza, B. M. P. S., Teixeira, C. M. M., & Guimarães, J. E. (2008). Perfil clínico-laboratorial da erliquiose monocítica canina em cães de Salvador e região metropolitana, Bahia. *Revista Brasileira Saúde Produção Animal*, 9(4), 770-776.
- Morais, H. A., Hoskins, J., Almosny, N. R. P., & Labarthe, N. V. (2004). Diretrizes gerais para diagnóstico e manejo de cães infectados por *Ehrlichia* spp. *Clínica Veterinária*, 9(48), 28-30.
- Moretti, L. D., Silva, A. V. da, Ribeiro, M. G., Paes, A. C., & Langoni, H. (2006). *Toxoplasma gondii* genotyping in a dog co-infected with distemper vírus and ehrlichiosis rickettsia. *Journal of the Institute of Tropical Medicine of São Paulo*, 6(48), 359-363. doi: 10.1590/S0036-46652006000600012
- Murphy, G. L., Ewing, S. A, Whitworth, L. C, Fox, J. C, & Kocan, A. A. (1998). A molecular and serologic survey of *Ehrlichia canis*, *E. chaffeensis*, and *E. ewingii* in dogs and ticks from Oklahoma. Veterinary *Parasitology*, 79, 325-339.
- Nakaghi, A. C. H., Machado, R. Z., Costa, M. T., André, M. R., & Baldani, C. D. (2008). Canine ehrlichiosis: clinical, hematological, serological and molecular aspects. *Ciência Rural*, 38(3), 766-700. doi: 10.1590/ S0103-84782008000300027
- Otranto, D., Dantas-Torres, F., & Breitschwerdt, E. B. (2009). Managing canine vector-borne diseases of zoonotic concern: parte one. *Trends in Parasitology*, *25*(4), 157-163. doi: 10.1016/j.pt2009.01.003
- Otranto, D., Caprariis, D. de, Lia, R. P., Tarallo, V., Lorusso, V., Testini, L., & Stanneck, D. (2010). Prevention of endemic canine vector-borne diseases using imidacloprid 10% and permethrin 50% in young dogs: a longitudinal field study. *Veterinary Parasitology*, *172*(3-4), 323-32. doi: 10.1016/j. vetpar.2010.05.017
- Perez, M., Bodor, M., Zhang, C., Xiong, Q., & Rikihisa, Y. (2006). Human infection with *Ehrlichia canis*

accompanied by clinical signs in Venezuela. *Annals New York Academy of Sciences*, *1078*(1), 110-117. doi: 10.1196/annals.1374.016

- Pritt, B. S., Allerdice, M. E. J., Sloan, L. M., Paddock, C. D., Munderloh, U. G., Rikihisa, Y., & Karpathy, S. E. (2017). Proposal to reclassify *Ehrlichia muris* as *Ehrlichia muris* subsp. *muris* subsp. nov. and description of *Ehrlichia muris* subsp. *eauclairensis* subsp. nov., a newly recognized tick-borne pathogen of humans. *International Journal of Systematic and Evolutionary Microbiology*, *67*(7), 2121-2126. doi: 10.1099/ijsem.0.001896
- Ribeiro, C. M., Matos, A. C., Azzolini, T., Bones, E. R., Wasnieski, E. A., Richinipereira, V. B, & Vidotto, O. (2017). Molecular epidemiology of *Anaplasma platys*, *Ehrlichia canis* and *Babesia vogeli* in stray dogs in Paraná, Brazil. *Pesquisa Veterinária Brasileira*, 37(2), 129-136. doi: 10.1590/S0100-736X2017000200006
- Saito, T. B. (2009). Estudo da erliquiose em cães expostos a carrapatos Rhipicephalus sanguineus experimentalmente infectados. Tese doutorado em Medicina Veterinária, Universidade de São Paulo, São Paulo, SP, Brasil. doi: 10.11606/T.10.2009.tde-20022009-134729
- Silva, G. C., Benitez A. N., Girotto, A., Taroda, A., Vidotto, M. C., Garcia, J. L., & Vidotto, O. (2012). Occurrence of *Ehrlichia canis* and *Anaplasma platys* in household dogs from northern Parana. *Revista Brasileira de Parasitologia Veterinária*, 21(4), 379-385. doi: 10.1590/S1984-29612012005000009
- Silva, I. P. M. (2015). Erliquiose canina revisão de literatura. *Revista Científica de Medicina Veterinária*, Ano XIII (24), 1-16.
- Sousa, V. R. F., Almeida, A. B. P. F., Barros, L. A., Sales, K. G., Justino, C. H. S., Dalcin, L., & Bomfim, T. C. B. (2010). Avaliação clínica e molecular de cães com erliquiose. *Ciência Rural*, 40(6), 1309-1313. doi: 10.1590/S0103-84782010000600011
- Trapp, S. M., Dagnone, A. S., Vidotto, O., Freire, R. L., Amude, A. M., & Morais, H. S. de. (2006). Soroepidemiologia da babesiose canina e erliquiose em população hospitalar. *Veterinary Parasitology*, *140*(3-4), 223-230. doi: 10.1016/j.vetpar.2006.03.030
- Ueno, T. E. H., Aguiar, D. M., Pacheco, R. C., Richtzenhain, J., Ribeiro, M. G., Paes, & Labruna, M. B. (2009). *Ehrlichia canis* em cães atendidos em hospital veterinário de Botucatu, Estado de São Paulo, Brasil. *Revista Brasileira de Parasitologia Veterinária*, 18(3), 57-61. doi: 10.4322/rbpv.01803010

- Vieira, R. F. C., Biondo, A. W., Guimarães, A. M. S., Santos, A. P., Santos, R. P., Dutra, L. H., & Vidotto, O. (2011). Ehrlichiosis in Brazil. *Revista Brasileira de Parasitologia Veterinária*, 20(1), 1-12. doi: 10.1590/S1984-29612011000100002
- Vieira, R. F. C., Vieira, T. S. W. J., Nascimento, D. A. G., Martins, T. F., Krawczak, F. S., Labruna, M. B., & Vidotto, O. (2013). Serological survey of *Ehrlichia* species in dogs, horses, and humans: Zoonotic scenery in a rural settlement from southern Brazil. *Revista do Instituto de Medicina Tropical*, 5(55), 335-40. doi: 10.1590/S0036-46652013000500007
- Witter, R., Vecchi, S. N., Pacheco, T. A., Melo, A. L. T., Borsa, A., Sinkoc, A. L., & Aguiar, D. M. (2013). Prevalência da erliquiose monocítica canina e anaplasmose trombocítica em cães suspeitos de hemoparasitose em Cuiabá, Mato Grosso. *Semina: Ciências Agrárias*, 34(6), p. 3811-3822. doi: 10.5433/1679-0359.2013v34n6Supl2p3811