

Physiological potential of *Physalis peruviana* L. seeds under different temperatures and light wavelengths

Potencial fisiológico de sementes de *Physalis peruviana* L. sob diferentes temperaturas e comprimentos de onda

Verônica Pellizzaro^{1*}; Mônica Satie Omura¹; Felipe Favoretto Furlan¹; Denis Santiago da Costa²; Raissa Marrafon Ponce¹; Gustavo Henrique Freiria¹; Helio Fernandes Ibanhes Neto¹; Lúcia Sadayo Assari Takahashi³

Resumo

Physalis peruviana L. é uma planta que produz pequenos frutos adocicados de grande importância nutricional e medicinal. Esse vegetal é propagado principalmente através de sementes, as quais necessitam de informações de respostas a temperatura e luz para germinação. Assim, o presente trabalho objetivou avaliar o uso de diferentes temperaturas e comprimentos de ondas de luz sobre o potencial fisiológico de lotes de sementes de *P. peruviana*. O experimento foi desenvolvido com sementes provenientes das safras 2015/15 e 2016/16. Os tratamentos foram: duas temperaturas de germinação (constante de 25°C e alternada de 20/30°C), dois lotes de sementes (safra 2015/15 e 2016/16) e quatro formas de exposições à luz durante a germinação (azul, vermelho, branca e escuro). Os dados foram submetidos a análise de componentes principais utilizando a matriz de correlação obtida por meio da média geral padronizada centrada em zero e variância 1 dos parâmetros avaliados para cada tratamento. Os autovalores e autovetores da matriz foram obtidos utilizando o software estatístico SAS University Edition[®]. As variáveis analisadas foram: Teste de germinação realizado no 7° e 28° dia considerando a protrusão de raiz, plântulas normais, comprimento de parte aérea, comprimento de raiz e massa de matéria seca total. Foi evidenciado que a germinação de sementes de *Physalis peruviana* L. deve ser realizada na presença de luz, nas temperaturas constante de 25°C ou alternada de 20/30°C. Quando submetidas ao teste no escuro, deve-se utilizar termoperíodo de 20-30 °C para proporcionar a expressão do máximo potencial fisiológico.

Palavras-chave: Fisális. Germinação. Termoperíodo.

Abstract

Physalis peruviana L. is a plant that produces small sweet berries of great nutritional and medicinal importance. This plant is propagated mainly through seeds, which requires information on germination response to temperature and light changes. Thus, the present study aimed to evaluate the use of different temperatures and light wavelengths on the physiological potential of *P. peruviana* seed batches. The experiment was conducted with seeds from plants cultivated in 2015/15 and 2016/16. The treatments

¹ Discentes, Universidade Estadual de Londrina, UEL, Londrina, PR, Brasil. E-mail: veronicapellizzaro@hotmail.com; monica_omura@hotmail.com; ffavorettofurlan@gmail.com; raissamp@hotmail.com; gustavo-freiria@hotmail.com; helioibanhesh@hotmail.com

² Prof., Instituto Federal de Educação, Ciência e Tecnologia de Mato Grosso do Sul, IFMS, Nova Andradina, MS, Brasil. E-mail: denisccauel@gmail.com

³ Prof^ª Dr^ª, Departamento de Agronomia, Centro de Ciências Agrárias, UEL, Londrina, PR, Brasil. E-mail: sadayo@uel.br

* Author for correspondence

applied were the following: two germination temperatures (constant temperature of 25 °C and alternated between 20/30 °C), two seed batches (2015/15 and 2016/16), and four forms of light exposure during germination (blue, red, white, and dark). The data were analyzed by principal component analysis using the correlation matrix obtained through the standardized mean equal to zero and variance 1 of the parameters evaluated for each treatment. The eigenvalues and eigenvectors of the matrix were obtained using the SAS University Edition® statistical software. The analyzed variables were: Germination test performed on the 7th and 28th day considering root protrusion, normal seedlings, shoot length, root length, and total dry matter mass. The data showed that the seed germination test of *Physalis peruviana* L. should be performed under a constant temperature of 25 °C, in the presence of light (white, blue, or red). When the option is the application of the test in the dark, it is necessary to use a thermoperiod of 20–30 °C to provide the expression of maximum physiological potential.

Key words: Goldenberry. Germination. Thermal periodicity.

Introduction

Physalis peruviana L. is a species belonging to the family of Solanaceae, widely recognized for having fruits with high amounts of vitamin C, vitamin A, vitamin B complexes, minerals, tocopherols, and carotenoids (RAMADAN, 2011). In addition to its nutritional importance, in Brazil the fruits are considered exotic, with high added value due to the flavor and the differential marketing of the fruits due to the presence of the calyx (LIMA et al., 2009).

In the process of agricultural production of this species, new plants are produced by means of asexual propagation, by using cuttings or *in vitro* cultivation, or by sexual multiplication, by using seeds (RUFATO et al., 2008). However, knowledge about its seed physiology is scarce compared to other solanaceous species, as there are no recommendations for germination of *P. peruviana* in the Rules for Seed Testing (BRASIL, 2009).

Temperature and light are two factors that influence the process of germination of the seeds. These parameters can be controlled, to optimize the speed, percentage, and synchronization of the germinative process, which results in reduced production expenses and more vigorous seedlings (NASSIF et al., 1998). Light and temperature requirements differ by species, since each one has its particular requirements in order for germination to occur quickly and uniformly (CARVALHO; NAKAGAWA, 2000).

Temperature variations have the potential to affect the percentage, the uniformity and the speed of germination of several species; and the optimum temperature is the one that permits the most efficient combination of speed and percentage of germination (MARCOS FILHO, 2015). According to the Rules for Seed Testing (BRASIL, 2009), for the species *Physalis alkekengi* and *Physalis pubescens*, the recommended temperature for the germination test is between 20–30 °C in the presence of light; however, most of the studies conducted with *Physalis peruviana* have been at a constant temperature of 25 °C, according to research by Fernandes et al. (2016), with X ray tests used for the evaluation of the physiological quality of the seeds of *Physalis peruviana* at different stages of development, and Souza et al. (2016), with the installation of germination tests in research carried out with seed storage of *P. peruviana*.

As for the light factor, some species have seeds that depend on the presence of light for germination, since germination is triggered by the formation of far-red phytochrome, which is formed by the absorption of light (red light corresponding to 660 nm); while others react indifferently to the presence of this factor (MARCOS FILHO, 2015). The phytochrome is a protein present in plant cells, which is associated with plant response to light stimuli. The influence of light on germination and the initial establishment of the plant has been the focus of several studies, leading to greater success in seedling production and an increase in

the establishment of plants in the initial phase of cultivation (MATOS et al., 2009; BANDEIRA et al., 2011; LOPES et al., 2005).

Bewley et al. (2013) point out that the response of seeds to light during germination is directly related to thermoperiodicity, as such, sensitivity varies by species. Heschel et al. (2007) report that germination depends on both phytochromes and temperature, and that even small changes in temperature have a great effect on the contribution of certain phytochromes to germination. Mondo et al. (2010) observe that different species had specific requirements for the germination of seeds, and that some species need light for twinning to occur.

Considering the above, the present study aimed to evaluate the effects of different temperatures and wavelengths of light on the physiological potential of seed batches of *P. peruviana*.

Material and Methods

Physalis peruviana L. seeds, represented by two batches from the Londrina State University production field, harvest 2015/15 and 2016/16, were extracted from fruits at full maturation stage (22 °C) and stored under refrigeration (10 °C) in a glass container with a polyethylene lid until the time of analysis. On the day prior to the application of the treatments, the seeds were removed from refrigeration and maintained at a temperature of 20 °C for complete thermal equilibrium.

For the evaluation of seed batch behavior, a principal component analysis was proposed, with four replicates, and the treatments were applied as described in Table 1. The two temperatures studied were proposed based on those most commonly used for other species of *Physalis*, available in the Rules for Seed Testing, since *P. peruviana* has no recommendation (BRASIL, 2009).

Table 1. Description of treatments such as temperature combinations, photoperiods and wavelengths for seed germination of *P. peruviana* seeds.

Wavelength ¹	Temperature ²	Lot
Blue - 440-485 nm	Constant (25 °C)	15/15
Blue - 440-485 nm	Constant (25 °C)	16/16
Blue - 440-485 nm	Alternating (20-30 °C)	15/15
Blue - 440-485 nm	Alternating (20-30 °C)	16/16
Red - 625-740 nm	Constant (25 °C)	15/15
Red - 625-740 nm	Constant (25 °C)	16/16
Red - 625-740 nm	Alternating (20-30 °C)	15/15
Red - 625-740 nm	Alternating (20-30 °C)	16/16
White ¹ - 370-750 nm	Constant (25 °C)	15/15
White - 370-750 nm	Constant (25 °C)	16/16
White - 370-750 nm	Alternating (20-30 °C)	15/15
White - 370-750 nm	Alternating (20-30 °C)	16/16
Dark	Constant (25 °C)	15/15
Dark	Constant (25 °C)	16/16
Dark	Alternating (20-30 °C)	15/15
Dark	Alternating (20-30 °C)	16/16

¹Wavelength relative to the visible light obtained by illumination with cold white lamps; ²Thermoperiod of 16 h at 30 °C and 8 h at 20 °C.

Initially, to avoid sensitization of the phytochromes in the seeds, sowing was carried out in an isolated room, which was surrounded by an external light whose internal illumination was coming from a green lamp, to which phytochrome is insensitive. Germination was obtained by equidistant distribution of 50 seeds per replicate, on blotter-type paper (10.5 × 10.5 cm), moistened with distilled water in an amount equivalent to 2.5 times the initial mass of the substrate, inside plastic boxes (11 × 11 × 3.5 cm). The plastic boxes (gerbox) were then wrapped in plastic film (cellophane) which allowed four forms of exposure to light (blue, red, white, and dark) and packed in transparent plastic bags to prevent loss of moisture.

The seed samples were taken to BOD-type chambers with either a constant temperature of 25 °C, under a 24h light regime, or alternated at 20–30 °C, with a light regime of 16 h at 30 °C and 8 h at 20 °C, as recommended for other species of *Physalis* in Rules for Seed Testing (BRASIL, 2009).

The protrusion of the primary root was evaluated at 7 and 28 days after sowing, and the number of normal seedlings at 28 days after sowing. The normality criterion was adopted, as proposed by the Rules for Seed Testing (BRASIL, 2009), in which seedlings that show potential to continue their development and give rise to normal plants when developed under favorable conditions are considered normal.

In addition, at the end of the germination test (28 days after sowing) the shoot and primary root length were determined by measuring the normal seedling parts, using a ruler graduated in centimeters adapted from Nakagawa (1999). Afterwards the parts were packed in kraft paper bags and kept in a forced circulation oven at 80 °C for 24 h, and then weighed on a precision scale adapted from Nakagawa (1999). The results were expressed in mg/seedling.

Principal component analysis was performed, using the correlation matrix obtained by using the standardized overall zero-centered mean and

variance 1 of the parameters evaluated for each treatment. This standardization was performed to avoid overestimating or underestimating the significance of a variable studied to the final result, due to differences in the scale of measurement of the parameters (MINGOTI; SILVA, 1997). The eigenvalues and eigenvectors of the matrix were obtained using the SAS statistical software (2000), University Edition®. The scores for each new component related to the treatments as well as the graphs were calculated and elaborated using Microsoft Office Excel software®.

Results and Discussion

According to the overall means presented in Table 2, differences in the seed behavior of *P. peruviana* under the proposed treatments can be observed. According to the principal components analysis, the parameters evaluated resulted in linear combinations giving rise to six new components, of which the first two were responsible for 81.52% of the total variation (Table 3). The first component, CP₁, presented the highest retention of data variability (58.82%), followed by the second component, CP₂ (22.70%), while the other components summed only retained a total of 18.48% of the variability. According to Jolliffe (2002), retention of the variability of the summed components must surpass 80%, for one to be able to interpret the results referring to the principal component analysis, in this case only two.

Analyzing the first component, represented by the linear equation $CP_1 = 0.475 \times (\text{Standardized PRP}) + 0.473 \times (\text{standardized FC}) + 0.472 \times (\text{standardized NS}) - 0.160 \times (\text{standardized APL}) + 0.396 \times (\text{standardized RL}) + 0.382 \times (\text{standardized TDM})$, it was observed that the primary root protrusion (PRP), first count (FC), and normal seed (NS), had the highest contributions and were considered the main characteristics in the construction of the new variable (0.475, 0.473 and 0.472, respectively). Thus, this component represented the germination

potential of the seeds of *P. peruviana*, and the higher the positive value of each treatment for this component, the higher the seedling performance. On the other hand, the second component, $CP_2 = 0.336 \times (\text{standardized PRP}) + 0.143 \times (\text{standardized FC}) + 0.366 \times (\text{standardized NS}) + 0.670 \times (\text{standardized APL}) - 0.407 \times (\text{standardized RL}) - 0.344 \times$

(standardized TDM), had the highest contributions (greater than 0.40 in module) provided by aerial part length (APL) and root length (RL) with 0.670 and -0.407 , respectively, and high positive values for this new component were related to better development of the seedlings during the test (Figure 1A).

Table 2. Mean of results for each treatment which were obtained for normal seedlings 28 days after sowing (NS), protrusion of the primary root 7 days after sowing (FC), primary root protrusion at 28 days after sowing (PRP), shoot length (APL), root length (RL) and total dry mass (TDM) for *P. peruviana*.

Wavelength	Temperature	Lot	NS (%)	FC (%)	PRP (%)	APL (mm)	RL (cm)	TDM (mg)
Blue ¹	Constant (25 °C)	15/15	92	42	95	30.1	1.57	4
Blue	Constant (25 °C)	16/16	27	38	46	17.8	2.56	4.25
Blue	Alternating (20-30 °C)	15/15	84	96	94	16.1	3.33	5
Blue	Alternating (20-30 °C)	16/16	39	24	47	11.2	2.27	2.25
Red ²	Constant (25 °C)	15/15	92	66	99	31.8	2.84	4.35
Red	Constant (25 °C)	16/16	56	34	64	27.3	2.42	4.25
Red	Alternating (20-30 °C)	15/15	90	96	99	13.1	3.33	4.25
Red	Alternating (20-30 °C)	16/16	25	35	38	5.5	1.62	1.45
White ³	Constant (25 °C)	15/15	100	58	100	31.0	2.83	4.9
White	Constant (25 °C)	16/16	45	26	50	20.1	3.54	5.85
White	Alternating (20-30 °C)	15/15	96	99	98	26.5	2.92	4.2
White	Alternating (20-30 °C)	16/16	38	44	40	4.5	2.23	5.85
Dark	Constant (25 °C)	15/15	8	3	8	52.7	1.52	1.5
Dark	Constant (25 °C)	16/16	14	0	16	39.0	0.55	2.25
Dark	Alternating (20-30 °C)	15/15	91	95	95	35.2	1.78	3.45
Dark	Alternating (20-30 °C)	16/16	74	42	91	31.9	103	2.85

¹Blue wavelength - 440-485 nm; ²Red wavelength - 625-740 nm; ³Visible wavelength - 370-750 nm.

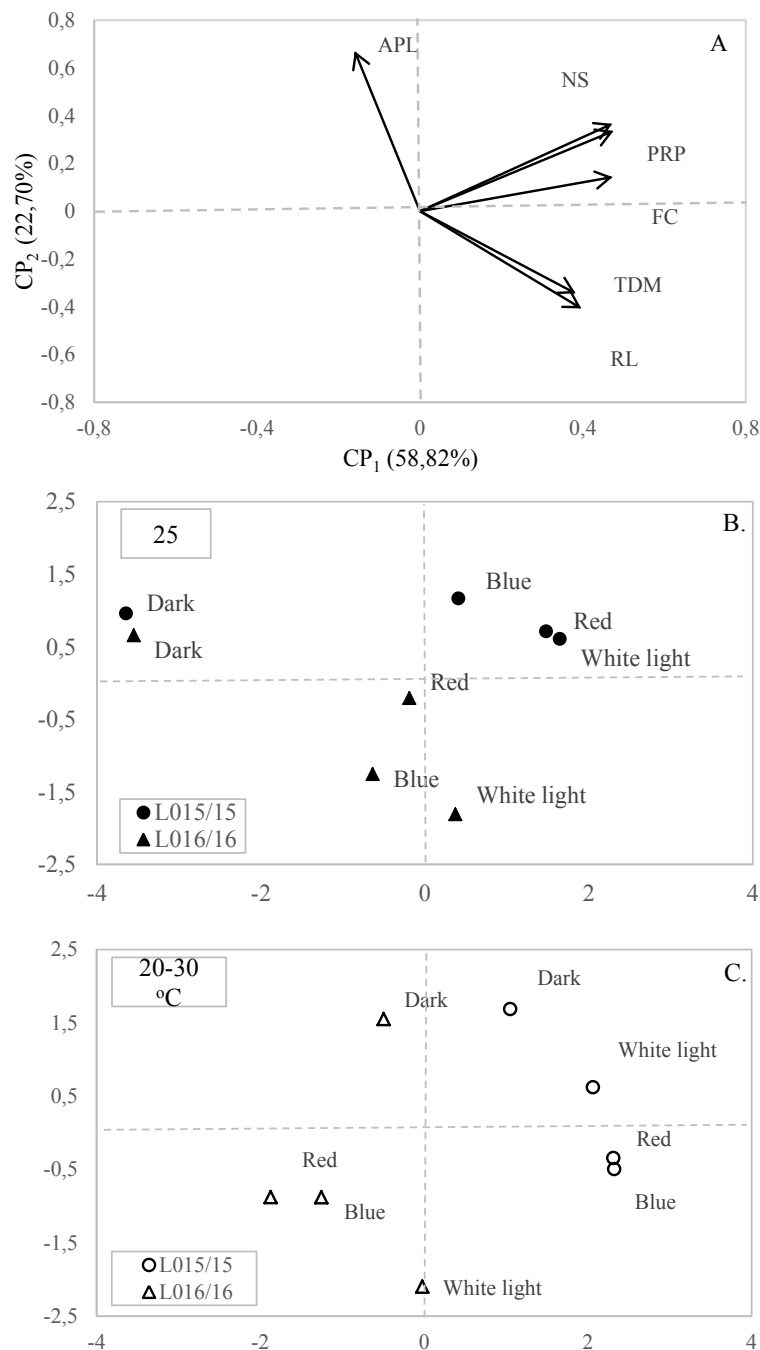
The results obtained for each wavelength at 25 °C (Figure 1B) led to the formation of three groups: the first group, with positive values for CP_1 and CP_2 (1st quadrant) was composed of the seeds in lot L015/15 that were germinated in the presence of white, red, and blue light. As previously described, CP_1 is related to the germination potential of the seeds, i.e., the higher the positive value for CP_1 the higher the seed performance in the presence of light (white, red or blue) at 25 °C for this batch (Figure 1B), results which are confirmed by Table 2.

The second group presented negative values for CP_1 and positive for CP_2 (2nd quadrant), and consisted of seeds that were germinated in the dark at 25 °C, with similar results for both L015/15 and L016/16 (Figure 1B). In this group, a low percentage of germination and seedlings were observed (Table 2). The third group was formed by the seeds of the lot L016/16, with negative values for CP_2 (3rd and 4th quadrants), germinated with wavelengths of white, red and blue light (Figure 1B). The values for CP_1 were intermediate, indicating an intermediate germinative potential (Table 2).

Thus, it was noted that at 25 °C, light was a determinant factor for seed germination of *P. peruviana*, since in the dark, germination occurred at very low rates for both batches studied. This may be related to seed dormancy and the need for light to germinate, because when the seeds were exposed to

blue, red or white light, germination rates were higher. According to Ozaslan et al. (2016), the germination of *Physalis angulata* and *Physalis philadelphica* is slightly affected by photoperiods, which suggests that the seeds are slightly photoblastic, indicating that the same behavior occurs in *P. peruviana*.

Figure 1 Results of principal component analysis. A - Eigenvectors for CP₁ and CP₂; B- Scores for wavelengths at 25 °C; C - Scores for wavelengths at 20–30 °C.



Light is commonly needed for germination in some species, as verified by Yamashita et al. (2011) in a study with *Conyza* seeds, in which light was a determining factor for germination. In an experiment conducted by Silva et al. (2016) with *Jatropha curcas* seeds, it was verified that in the presence of light, there is a homogenization of the germinative

parameters in all batches analyzed, which does not happen in the dark. Under controlled conditions, Cardoso et al. (2015) verified that a temperature of 20 °C, the presence of constant light and a filter paper substrate led to the most uniform germination of *P. peruviana* seeds.

Table 3. Results obtained from the correlation matrix of the results of primary root protrusion (PRP), first count (FC), normal seedlings (NS), shoot length (APL) root length (RL), and total dry mass (TDM) to *P. peruviana*.

ComponentsMatrix eigenvalues.....			
	Eigenvalue	Difference	Proportion	Cumulative
CP ₁	3.5295	2.1676	58.82 %	58.82 %
CP ₂	1.3618	0.7292	22.70 %	81.52 %
CP ₃	0.6326	0.3222	10.54 %	92.07 %
CP ₄	0.3104	0.1564	5.17 %	97.24 %
CP ₅	0.1540	0.1423	2.57 %	99.81 %
CP ₆	0.0116	-	0.19 %	100.00 %

The first component efficiently separated the seeds from lot L015/15 of L016/16, by analyzing the results obtained under alternating temperatures of 20-30 °C (Figure 1C), forming two distinct groups (1st and 4th quadrant for L015/15; 3rd and 4th quadrant for L016/16). However, there was no separation for the seeds that were germinated in the dark, contrary to the result obtained at 25 °C (Figure 1B). It was thus observed that the thermoperiodicity provided by alternating 20-30 °C may be a solution for overcoming seed dormancy in *P. peruviana* (Table 2). The influence of the far-red phytochrome on the temperature required for overcoming dormancy may be associated with the effects of temperature on processes mediated by the phytochrome (PONS, 2000). According to Heschel et al. (2007), small changes in temperature have a large effect on the contribution of particular phytochromes to germination.

According to Takaki (2001), seeds of *P. peruviana* germinate both in the presence of light and in the dark, and can be classified as neutral photoblastic. However, this categorization cannot

be considered definitive, since other factors, such as humidity and temperature, can alter their photoblastic characteristics, a fact confirmed by the results obtained for germination in the dark under thermoperiodicity (Table 2 and Figure 1C).

Regarding the batches, although different values were observed between the two harvests due to the lower physiological quality of L016/16, the response was similar for the temperature and light conditions, indicating that their behavior is independent of the initial seed quality. Thus, it was verified that for the germination physiology of *P. peruviana* seeds, the light and temperature factors strongly interact with each other, and are able to influence principally the number of normal seedlings germinated and seedling length.

Conclusions

The seed germination test of *Physalis peruviana* L. must be performed under a constant temperature of 25 °C, in the presence of light (white, blue or red). When the test is performed in the dark, the

thermoperiod of 20–30 °C should be used to provide the expression of maximum physiological potential.

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